

# polySegratioMM: An R library for Bayesian mixture models for marker dosage in autopolyploids

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It is well known that the dosage level of markers in autopolyploids and allopolyploids can be characterised by their observed segregation ratios. The related package **polySegratio** provides functions to allocate dosage by standard approaches and to simulate marker data sets for differing ploidies and levels of overdispersion. Note that these methods could equally well be applied to allopolyploids with specified expected segregation ratios. For details see **polySegratio**.

A Bayesian approach to marker dosage estimation was proposed by Baker et al SUBMITTED whereby all markers are fitting a finite mixture distribution.

This library calls the **JAGS** software for Bayesian calculation. **JAGS 1.0** or higher must be installed following instructions from <http://www-fis.iarc.fr/~martyn/software/jags/>. **JAGS** must be installed and the executable must be in your path. Note that no checking is carried out to ascertain whether or not **JAGS** is set up appropriately.

To use the library, you need to attach it with

```
> library(polySegratioMM)
```

## 1 Simulated data

Library functions are demonstrated on a simulated data set generated using the **sim.autoMarkers** function from the **polySegratio** package.

The following R code can be used to generate 500 markers for 200 auto-hexaploid individuals exhibiting overdispersion with the parameter **shape1** = 25. The underlying percentages of single double and triple dose markers are 70%, 20% and 10%, respectively.

```
hexmarkers <- sim.autoMarkers(6,c(0.7,0.2,0.1),n.markers=500,n.individuals=200)

> print(hexmarkers)
```

Autopolyploid dominant markers generated at Fri Jul 18 14:51:38 2008  
with call:

```
sim.autoMarkers(ploidy.level = 6, dose.proportion = c(0.7, 0.2,
```

```
0.1), n.markers = 500, n.individuals = 200)
```

Ploidy level is: 6 ( Hexaploid )

Parents were set as heterogeneous for the markers

Theoretical segregation proportions:

ratio.SD	ratio.DD	ratio.TD	ploidy.level
"0.5"	"0.8"	"0.95"	"6"
ploidy.name	type.parents		
"Hexaploid"	"heterogeneous"		

Proportions in each dosage class:

SD	DD	TD
0.7	0.2	0.1

No. of markers generated from multinomial distribution:

No.markers	
SD	346
DD	103
TD	51

Data were generated for 200 individuals with 500 markers

A subset is:

	X.1	X.2	X.3	X.4	X.5	X.6	X.7	X.8	X.9	X.10	r	n	ratio	dose
M.1	1	0	0	1	0	1	0	1	1	1	108	200	0.54	SD
M.2	0	1	1	1	0	1	0	1	0	1	102	200	0.51	SD
M.3	1	0	0	1	0	0	0	1	1	1	103	200	0.515	SD
M.4	1	1	1	1	1	0	0	0	1	0	97	200	0.485	SD
M.5	0	0	1	1	0	0	1	1	0	1	99	200	0.495	SD
M.6	1	1	0	0	0	0	0	0	0	0	103	200	0.515	SD
M.7	1	1	0	0	0	1	0	0	1	1	101	200	0.505	SD
M.8	1	1	1	1	1	1	0	0	0	1	102	200	0.51	SD
M.9	0	1	1	1	1	0	0	1	1	0	110	200	0.55	SD
M.10	1	0	1	0	1	1	1	0	1	1	108	200	0.54	SD

Note that the segregation ratios for simulated or real data may be extracted by using `segregationRatios` which sets up the appropriate objects for testing marker dosage and plotting or summarising the marker data.

```
> sr <- segregationRatios(hexmarkers$markers)
```

For instance, as seen in Figure 1, segregation ratios may be plotted with

```
plotTheoretical(ploidy.level=6, seg.ratios=sr,
  expected.segratio=NULL, proportions=c(0.7,0.2,0.1),
  n.individuals=200)
```

On the other hand, consider a similar data set that exhibits overdispersion. This may be simulated as follows

```
hexmarkers.overdisp <- sim.autoMarkers(6,c(0.7,0.2,0.1),n.markers=500,n.individuals=200,
  overdispersion=TRUE, shape1=30)
```

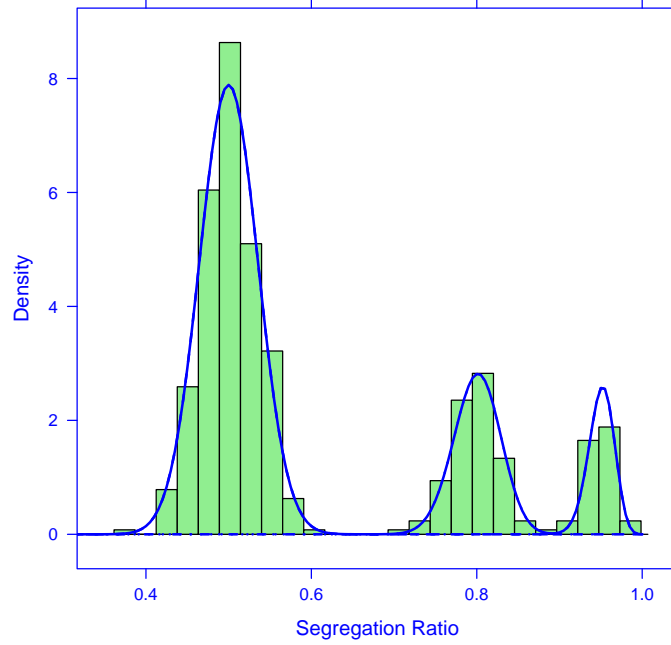


Figure 1: Segregation ratios of 500 simulated markers from 200 autohexaploid individuals. Percentages of single double and triple dose markers are 70%, 20% and 10%, respectively. Data were generated assuming no overdispersion.

```
> sr.overdisp <- segregationRatios(hexmarkers.overdisp$markers)
```

The histogram of marker segregation ratios, which is a useful graphical method for identifying overdispersion or outliers, is seen in Figure 2. Note that, due to overdispersion the theoretical distribution is narrow than the observed data.

## 2 A Bayesian mixture model approach

For the  $j^{th}$  marker  $j = 1 \dots n$ , we assume the observed number  $r_j$  of dominant markers out of  $N_j$  lines follows a binomial distribution denoted  $Bin(N_j, P_k)$ . If we knew the dosage  $k$  then, following Ripol et al. (1999), the expected value of  $P_k$  may be written as

$$P_k(k|m, x) = 1 - \frac{\binom{m-k}{mx}}{\binom{m}{mx}}, k = 0 \dots m/2 \quad (1)$$

where  $m$  is the ploidy level or number of homologous chromosomes and the monoploid number  $x$  is the number of chromosomes in a basic set. Note that

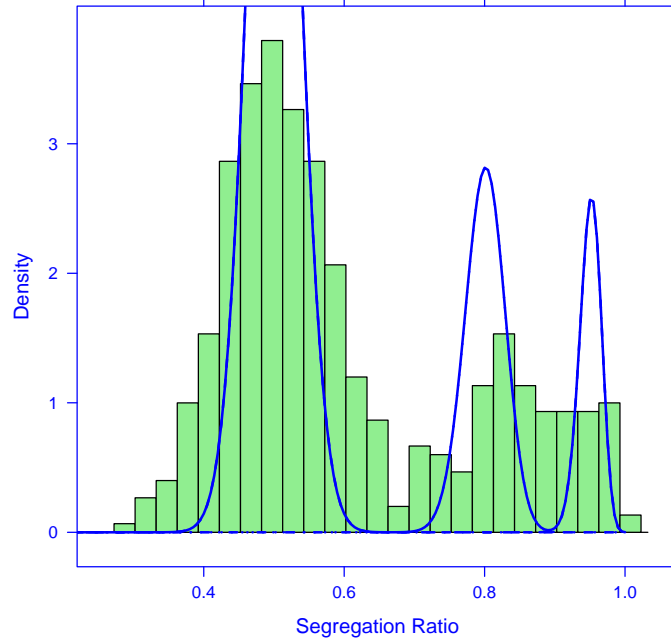


Figure 2: Segregation ratios of 500 simulated markers from 200 autohexaploid individuals. Percentages of single double and triple dose markers are 70%, 20% and 10%, respectively. Data were generated from the Beta-Binomial distribution assuming a shape parameter `shape1` of 30.

for diploids  $m = 2$ , tetraploids  $m = 4$ , octaploids then  $m = 8$  and so on and also that if there are no marker data missing then  $N_j$  is simply the number of progeny.

Since the dosage of each marker is unknown, we rely on the missing data representation of Dempster et al. (1977) and Tanner and Wong (1987) which is commonly adopted for MCMC computation in finite mixture models. An indicator variable  $z_j$  corresponding to unknown marker dosage class  $k$  is introduced where  $z_j = k$  if the marker has dose  $k$ . For the  $K$  components with  $K \leq m/2$ , consider the logit transformation of the true segregation proportions  $P_k$  for dose  $k, k = 1 \dots K$ . The the logit transformed segregation ratio  $\omega_k$  is then

$$\omega_k = \log\left(\frac{P_k}{1 - P_k}\right). \quad (2)$$

Let  $z = (z_1 \dots z_n)^T$  be a vector of unknown dosages (labelled  $1, 2 \dots K$  corresponding to simplex, duplex, triplex markers and so on), then  $r_j$  is binomially distributed with known size parameter  $N_j$  and unknown proportion parameter  $\omega_{z_j}$  which is the segregation ratio for marker dosage  $z_j$ . Hence, given marker

dosage  $z_j$  then

$$r_j|z_j \sim \text{Bin}(N_j, \omega_{z_j}), \quad (3)$$

where

$$\text{logit}(\omega_{z_j}) = \log\left(\frac{\omega_{z_j}}{1 - \omega_{z_j}}\right) \sim N(\mu_{z_j}, \tau_{z_j}^{-1})$$

where  $\mu_k$  and  $\tau_k$  are the mean and precision ( $\tau_k = 1/\sigma_k^2$ ) of marker dosage class  $k$  on the logit scale.

Since the dosage is unknown, for the autohexaploid data generated here then for the  $\text{logit}(\omega_{z_k})$  can be modelled as a finite mixture of 3 normals

$$\text{logit}(\omega_{z_j}) \sim \pi_1 N(\mu_1, \tau_1^{-1}) + \pi_2 N(\mu_2, \tau_2^{-1}) + \dots + \pi_K N(\mu_K, \tau_K^{-1}) \quad (4)$$

where  $\mu_k$  is the mean and  $\tau_k$  is the precision of component  $k$  on the logit scale, and  $\pi_k$  are the mixing proportions of the three components with  $\sum_{k=1}^K \pi_k = 1$ . The probability density function  $f(x)$  of  $\text{logit}(\omega_k)$  is

$$f(x) = \sum_{k=1}^K \pi_k \phi(x|\mu_k, \tau_k^{-1}) \quad (5)$$

where  $\phi$  is the normal cumulative distribution function with parameters mean  $\mu_k$  and variance  $\sigma_k^2 = \tau_k^{-1}$ .

Simulation studies suggested that incorporating strong prior information, such as the expected distributions of Haldane (1930) provided the best method of allocating dosage. Further details may be found in Baker, et al SUBMITTED

### 3 Specifying a model

A mixture model may be set up with `setModel`. By default, only two parameters are required, namely the `ploidy.level` or the number of homologous chromosomes set either as a numeric or as a character string and also `n.components` or the number of components for mixture model (less than or equal to maximum number of possible dosages). By default, strong priors are set by using the formulae of Haldane (1930) for the expected numbers and ratios of offspring for various parental configurations of autopolyploids.

For the autohexaploid data generated above, the models are set with

```
> x.mod1 <- setModel(3, 6)
```

The R object `x.mod1` contains components describing aspects of the model such as the number of components, ploidy, expected segregation ratios and so on. Note that the `str` command is useful for displaying the internal structure of any R object.

### 4 Fitting a mixture model

While various options are available for fine tuning the MCMC process, the simplest way to fit a mixture model to allocate marker dosages is with the wrapper function `runSegratioMM` as follows:

```
mcmcHexRun <- runSegratioMM(sr.overdisp, x.mod1)
```

which automatically determines starting values, priors, length of burn in, number of iterations, and other parameters as well as producing summary statistics and diagnostic plots.

To run JAGS without producing plots then set the `plots` option to `FALSE`. For the overdispersed data running this command produced the following selected output. While selected output is printed here the simple command `print(mcmcHexRun)` would produce the following output and more.

The summary of processing times:

```
> print(mcmcHexRun$run.jags)
```

```
CMD File: test.cmd
```

```
JAGS started at Fri Jul 18 14:51:41 2008
```

```
JAGS run completed successfully at Fri Jul 18 14:55:47 2008
```

```
Elapsed times:
```

```
  user  system elapsed
228.0   228.0   246.4
```

And summary statistics for the posterior distributions of selected parameters:

```
> print(mcmcHexRun$summary)
```

```
$statistics
```

	Mean	SD	Naive SE	Time-series SE
P[1]	0.72356	0.02064	0.0002919	0.0003616
P[2]	0.19709	0.01906	0.0002695	0.0005650
P[3]	0.07935	0.01336	0.0001890	0.0006636
mu[1]	0.02381	0.01696	0.0002398	0.0006163
mu[2]	1.54959	0.04421	0.0006253	0.0031491
mu[3]	3.16008	0.08879	0.0012557	0.0074792
sigma	0.27012	0.01215	0.0001718	0.0005203

```
$quantiles
```

	2.5%	25%	50%	75%	97.5%
P[1]	0.681698	0.70954	0.72439	0.73818	0.76208
P[2]	0.161445	0.18404	0.19643	0.20946	0.23651
P[3]	0.054776	0.07029	0.07876	0.08805	0.10753
mu[1]	-0.008683	0.01177	0.02381	0.03529	0.05805
mu[2]	1.464629	1.51904	1.54875	1.57957	1.63693
mu[3]	2.995540	3.09972	3.15792	3.21625	3.34191
sigma	0.247668	0.26165	0.26998	0.27826	0.29443

```
$start
```

```
[1] 0
```

```
$end
```

```
[1] 4999
```

```
$thin
```

```
[1] 1
```

```
$nchain
```

```
[1] 1
```

```
attr("class")
```

```
[1] "summarySegratioMCMC"
```

Note that MCMC convergence diagnostic output is produced automatically. Assessing convergence is crucial in MCMC and poor convergence may result in mis-allocated marker dosages. The diagnostic statistics indicate that convergence was achieved.

```
> print(mcmcHexRun$diagnostics)
```

```
$raftery
```

```
$raftery[[1]]
```

```
Quantile (q) = 0.025
```

```
Accuracy (r) = +/- 0.005
```

```
Probability (s) = 0.95
```

	Burn-in (M)	Total (N)	Lower bound (Nmin)	Dependence factor (I)
P[1]	2	3803	3746	1.020
P[2]	2	3930	3746	1.050
P[3]	2	3680	3746	0.982
mu[1]	4	4713	3746	1.260
mu[2]	10	11010	3746	2.940
mu[3]	18	19611	3746	5.240
sigma	10	10754	3746	2.870

```
$geweke
```

```
$geweke[[1]]
```

```
Fraction in 1st window = 0.1
```

```
Fraction in 2nd window = 0.5
```

P[1]	P[2]	P[3]	mu[1]	mu[2]	mu[3]	sigma
1.3314	-1.9552	1.4159	1.7912	-0.7923	-0.9007	-1.0033

```
$heidel
```

```
$heidel[[1]]
```

	Stationarity test	start iteration	p-value
P[1]	passed	1	0.348
P[2]	passed	1	0.904
P[3]	passed	1	0.547
mu[1]	passed	1	0.387
mu[2]	passed	1	0.465
mu[3]	passed	1	0.374
sigma	passed	1	0.913

	Halfwidth test	Mean	Halfwidth
P[1]	passed	0.7236	0.000709
P[2]	passed	0.1971	0.001107
P[3]	passed	0.0794	0.001301
mu[1]	passed	0.0238	0.001208
mu[2]	passed	1.5496	0.006172
mu[3]	passed	3.1601	0.014659
sigma	passed	0.2701	0.001020

```
$hpd
```

```
$hpd[[1]]
```

	lower	upper
P[1]	0.68391	0.76365
P[2]	0.15950	0.23382
P[3]	0.05438	0.10695
mu[1]	-0.01018	0.05593
mu[2]	1.46539	1.63737
mu[3]	2.99198	3.33813
sigma	0.24697	0.29359

```
attr("Probability")
[1] 0.95
```

And finally, summaries of marker dosage allocations are produced:

```
> print(mcmcHexRun$doses)
```

```
Dosages for chain: 1
```

```
Thresholds set at:
```

```
[1] 0.50 0.60 0.70 0.80 0.90 0.95 0.99
```

```
A random sample of posterior probabilities and classifications
```

	SD	DD	TD	0.5	0.6	0.7	0.8	0.9	0.95	0.99	maxPostP
M.5	1.0000	0.0000	0.0000	1	1	1	1	1	1	1	1
M.8	1.0000	0.0000	0.0000	1	1	1	1	1	1	1	1
M.77	1.0000	0.0000	0.0000	1	1	1	1	1	1	1	1
M.90	1.0000	0.0000	0.0000	1	1	1	1	1	1	1	1



M.154	0.9998	0.0002	0.0000	1	1	1	1	1	1	1	1
M.161	1.0000	0.0000	0.0000	1	1	1	1	1	1	1	1
M.172	1.0000	0.0000	0.0000	1	1	1	1	1	1	1	1
M.184	0.9998	0.0002	0.0000	1	1	1	1	1	1	1	1
M.226	1.0000	0.0000	0.0000	1	1	1	1	1	1	1	1
M.231	1.0000	0.0000	0.0000	1	1	1	1	1	1	1	1
M.275	1.0000	0.0000	0.0000	1	1	1	1	1	1	1	1
M.297	1.0000	0.0000	0.0000	1	1	1	1	1	1	1	1
M.312	1.0000	0.0000	0.0000	1	1	1	1	1	1	1	1
M.318	1.0000	0.0000	0.0000	1	1	1	1	1	1	1	1
M.328	1.0000	0.0000	0.0000	1	1	1	1	1	1	1	1
M.347	1.0000	0.0000	0.0000	1	1	1	1	1	1	1	1
M.407	0.0000	0.9996	0.0004	2	2	2	2	2	2	2	2
M.447	0.0000	0.9972	0.0028	2	2	2	2	2	2	2	2
M.450	0.0000	1.0000	0.0000	2	2	2	2	2	2	2	2
M.461	0.0000	0.4230	0.5770	3	.	.	.	.	.	.	3

Maximum posterior probabilities for 500 markers

Min.	1st Qu.	Median	Mean	3rd Qu.	Max.
0.502	1.000	1.000	0.980	1.000	1.000

Proportion of genes classified using maximum posterior probability

SD	DD	TD
0.724	0.194	0.082

Total proportion of markers classified: 1

Call:

```
dosagesJagsMix(mcmc.mixture = read.jags, jags.control = jags.control,
  seg.ratio = seg.ratios)
```

Note that simply plotting `mcmcHexRun` will produce a histogram of segregation proportions and the fitted model but that other plots are easily produced.

When `plots` option of `runSegratioMM` is set to the default value of `TRUE`, numerous plots are produced including trace and density plots from the `CODA` package. These may also be extracted manually but the process is somewhat more complicated. For instance to obtain trace and density plots for the parameters  $p_1$ ,  $\mu_1$ ,  $\sigma_1$  and for the 140<sup>th</sup> marker, as shown in Figure 3, then `CODA` may be used directly by following command.

```
plot(mcmcHexRun$mcmc.mixture$mcmc.list[[1]][,c("P[1]", "mu[1]", "sigma", "T[140]")])
```

The histogram of segregation proportions with fitted and theoretical values shown in Figure 4 may be obtained by setting the `theoretical` option to `TRUE` as follows.

```
print(plot(mcmcHexRun, theoretical=TRUE))
```

NULL

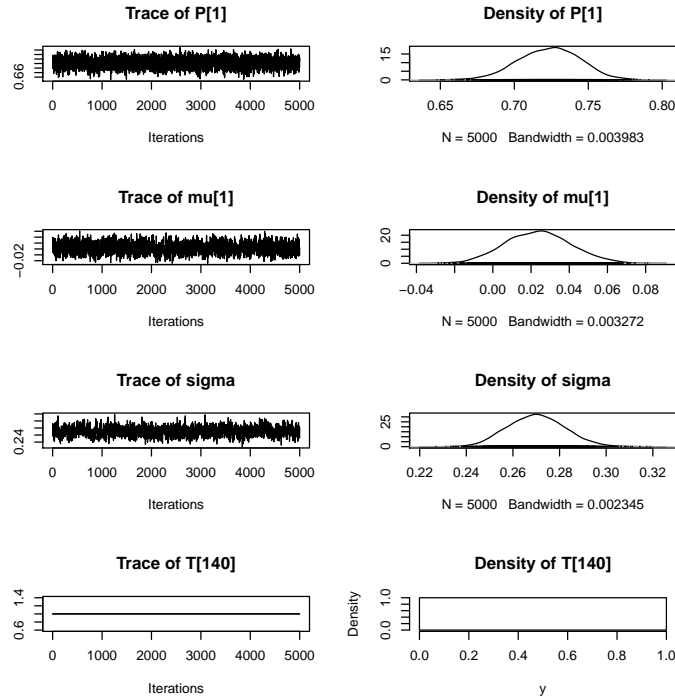


Figure 3: Trace and posterior density plots for the parameters  $p_1$ ,  $\mu_1$ ,  $\sigma_1$  and for the 140<sup>th</sup> marker for the overdispersed data.

## 5 Assigning marker dosage

Marker dosages allocations may be obtained directly from the object `mcmcHexRun`. The dosage with maximum posterior probability is simply `mcmcHexRun$doses$max.post.dosage`. A more conservative allocation is obtained by using `mcmcHexRun$doses$dosage[, "0.8"]` whereby the dosage with posterior probability over 0.8 is employed. For instance, to tabulate the number of markers (including those not allocated a dosage which are labelled NA) the `table` command can be employed.

```
> cat("Employing maximum posterior probability\n")
```

```
Employing maximum posterior probability
```

```
> table(Dose = mcmcHexRun$doses$max.post.dosage, exclude = NULL)
```

```
Dose
  1    2    3 <NA>
362  97  41    0
```

```
> cat("Employing posterior probability > 0.8\n")
```

Warning: component proportions normalised, now:

```

P[1]    P[2]    P[3]
0.72356 0.19709 0.07935

```

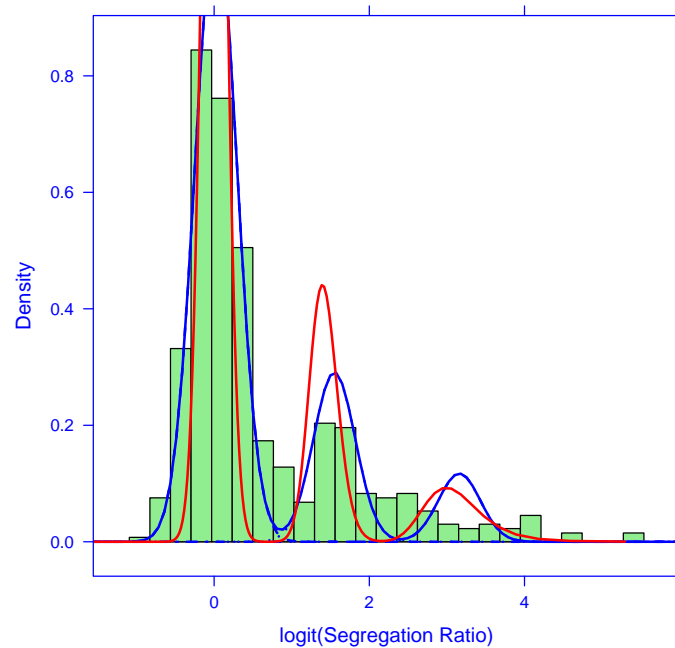


Figure 4: Fitted (blue) and theoretical (red) distributions for simulated segregation ratios with overdispersion for 500 markers from 200 individuals.

Employing posterior probability > 0.8

```
> table(Dose = mcmcHexRun$doses$dosage[, "0.8"], exclude = NULL)
```

```

Dose
  1    2    3 <NA>
358  89  34   19

```

And of course since the data were simulated we can compare the estimated and true dosages obtained as `hexmarkers.overdisp$true.doses$dosage` via cross tabulation. Doses can also be obtained for the standard  $\chi^2$  test by using the `test.segRatio` command from the `polySegratio` library.

```
> cat("Employing theChi squared test\n")
```

Employing theChi squared test

```

> dose.chi <- test.segRatio(sr.overdisp, ploidy.level = 6)
> table(Chi2Dose = dose.chi$dosage, True = hexmarkers.overdisp$true.doses$dosage,
+       exclude = NULL)

```

```

      True
Chi2Dose  1   2   3 <NA>
  1    223   2   0   0
  2     0  54   5   0
  3     0   3  27   0
<NA> 130  39  17   0

> cat("Employing maximum posterior probability\n")

Employing maximum posterior probability

> table(MixtureDose = mcmcHexRun$doses$max.post.dosage,
+       True = hexmarkers.overdisp$true.doses$dosage, exclude = NULL)

      True
MixtureDose  1   2   3 <NA>
  1    353   9   0   0
  2     0  86  11   0
  3     0   3  38   0
<NA>    0   0   0   0

> cat("Employing posterior probability > 0.8\n")

Employing posterior probability > 0.8

> table(MixtureDose = mcmcHexRun$doses$dosage[, "0.8"],
+       True = hexmarkers.overdisp$true.doses$dosage, exclude = NULL)

      True
MixtureDose  1   2   3 <NA>
  1    352   6   0   0
  2     0  78  11   0
  3     0   2  32   0
<NA>    1  12   6   0

```

These tables show that far fewer markers are allocated a dosage using the standard  $\chi^2$  test than by the mixture model. Fewer markers were misclassified using a posterior probability threshold of 0.8 rather than the maximum posterior probability as a basis for allocating dosage.

## References

- Dempster, A. P., Laird, N. M., and Rubin, D. B. (1977). Maximum likelihood from incomplete data via the EM algorithm (with discussion). *Journal of the Royal Statistical Society B*, 39:1–38.
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- Ripol, M. I., Churchill, G. A., da Silva, J. A., and Sorrells, M. (1999). Statistical aspects of genetic mapping in autopolyploids. *Gene*, 235(1-2):31–41.

Tanner, M. A. and Wong, W. H. (1987). The calculation of posterior distributions by data augmentation: with discussion. *Journal of the American Statistical Association*, 82:528–550.

## 5.1 Acknowledgments

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