

Package ‘lme4GS’

July 22, 2025

Type Package

Title 'lme4' for Genomic Selection

Version 0.1

Date 2025-04-06

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Depends R (>= 3.6.3), lme4, methods, Matrix

Suggests BGLR

Description Flexible functions that use 'lme4' as computational engine for fitting models used in Genomic Selection (GS). GS is a technology used for genetic improvement, and it has many advantages over phenotype-based selection. There are several statistical models that adequately approach the statistical challenges in GS, such as in linear mixed models (LMMs). The 'lme4' is the standard package for fitting linear and generalized LMMs in the R-package, but its use for genetic analysis is limited because it does not allow the correlation between individuals or groups of individuals to be defined. The 'lme4GS' package is focused on fitting LMMs with covariance structures defined by the user, bandwidth selection, and genomic prediction. The new package is focused on genomic prediction of the models used in GS and can fit LMMs using different variance-covariance matrices. Several examples of GS models are presented using this package as well as the analysis using real data. For more details see Caamal-Pat et.al. (2021) <[doi:10.3389/fgene.2021.680569](https://doi.org/10.3389/fgene.2021.680569)>.

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NeedsCompilation no

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Repository CRAN

Date/Publication 2025-04-08 15:30:02 UTC

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lme4GS-package	<i>lme4 for Genomic Selection</i>
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Description

Flexible functions that use 'lme4' as computational engine for fitting models used in Genomic Selection (GS). GS is a technology used for genetic improvement, and it has many advantages over phenotype-based selection. There are several statistical models that adequately approach the statistical challenges in GS, such as in linear mixed models (LMMs). The 'lme4' is the standard package for fitting linear and generalized LMMs in the R-package, but its use for genetic analysis is limited because it does not allow the correlation between individuals or groups of individuals to be defined. The 'lme4GS' package is focused on fitting LMMs with covariance structures defined by the user, bandwidth selection, and genomic prediction. The new package is focused on genomic prediction of the models used in GS and can fit LMMs using different variance-covariance matrices. Several examples of GS models are presented using this package as well as the analysis using real data. For more details see Caamal-Pat et.al. (2021) <doi:10.3389/fgene.2021.680569>.

lmerUvcov	<i>Fits a linear mixed model with user specified variance covariance-matrices.</i>
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Description

Fits a linear mixed model with user specified variance covariance-matrices.

Usage

```
lmerUvcov(formula, data = NULL, Uvcov = NULL, verbose=0L)
```

Arguments

formula	a two-sided linear formula object describing both the fixed-effects and random-effects part of the model, with the response on the left of a '~' operator and the terms, separated by '+' operators, on the right. Random-effects terms are distinguished by vertical bars ' ' separating expressions for design matrices from grouping factors.
data	an optional data frame containing the variables named in 'formula'.
Uvcov	list.
verbose	integer scalar, verbose output from optimizeLmer function?. If '> 0' verbose output is generated during the optimization of the parameter estimates, default value is 0L.

Details

The routine fits the linear mixed model:

$$\mathbf{y} = \mathbf{X}\boldsymbol{\beta} + \mathbf{Z}_1\mathbf{u}_1 + \cdots + \mathbf{Z}_q\mathbf{u}_q + \mathbf{e},$$

where $\mathbf{y}$ is the response vector, $\mathbf{X}$ is the matrix for fixed effects, $\boldsymbol{\beta}$ is the vector of fixed effects, $\mathbf{Z}_j$ is a design matrix for random effects, $\mathbf{u}_j$ is a vector of random effects, $j = 1, \dots, q$. We assume that $\mathbf{u}_j \sim N(\mathbf{0}, \sigma_j^2 \mathbf{K}_j)$, $j = 1, \dots, q$ and $\mathbf{e} \sim N(\mathbf{0}, \sigma_e^2 \mathbf{I})$.

The linear mixed model can be re-written as:

$$\mathbf{y} = \mathbf{X}\boldsymbol{\beta} + \mathbf{Z}_1^*\mathbf{u}_1^* + \cdots + \mathbf{Z}_q^*\mathbf{u}_q^* + \mathbf{e},$$

where $\mathbf{Z}_j^* = \mathbf{Z}_j \times \mathbf{L}_j$, with $\mathbf{L}_j$ from Cholesky factorization for $\mathbf{K}_j$. Alternatively, $\mathbf{Z}_j^* = \mathbf{Z}_j \times \boldsymbol{\Gamma}_j \boldsymbol{\Lambda}_j^{1/2}$, with $\boldsymbol{\Gamma}_j$ and $\boldsymbol{\Lambda}_j$ the matrix of eigen-vectors and eigen-values obtained from the eigen-value decomposition for $\mathbf{K}_j$. The factorization method for $\mathbf{K}_j$ is selected automatically at runtime.

Value

An object of class merMod (more specifically, an object of subclass lmerMod), for which many methods are available (e.g. methods(class="merMod"))

Author(s)

Paulino Perez-Rodriguez

References

Caamal-Pat D., P. Perez-Rodriguez, J. Crossa, C. Velasco-Cruz, S. Perez-Elizalde, M. Vazquez-Pena. 2021. lme4GS: An R-Package for Genomic Selection. *Front. Genet.* **12**:680569. doi: 10.3389/fgene.2021.680569 doi: 10.3389/fgene.2021.680569

Examples

```
library(BGLR)
library(lme4GS)

#####
#Example wheat
#####
data(wheat)
X<-wheat.X
Z<-scale(X,center=TRUE,scale=TRUE)
G<-tcrossprod(Z)/ncol(Z)
A<-wheat.A
rownames(G)<-colnames(G)<-rownames(A)
y<-wheat.Y[,1]

data<-data.frame(y=y,m_id=rownames(G),a_id=rownames(A))

fm1<-lmerUvcov(y~(1|m_id)+(1|a_id),data=data,
               Uvcov=list(m_id=list(K=G),a_id=list(K=A)))

summary(fm1)

#Predictions
plot(y,predict(fm1))

#Random effects
ranef(fm1)
```

lmerUvcov-class

User defined variance covariance mixed-effects model fits

Description

A mixed-effects model fit by [lmerUvcov](#). This class extends class "[merMod](#)" class and includes one additional slot, `relfac`, which is a list of (left) Cholesky factors of the relationship matrices derived from user provided variance covariance matrixes.

Objects from the Class

Objects are created by calls to the [lmerUvcov](#) function.

Extends

Class "[merMod](#)", directly.

Methods

ranef signature(object = "lmerUvcov"): incorporates the user defined variance covariance matrix into the random effects as returned for the object viewed as a "merMod" object.

See Also

[lmerUvcov](#)

Examples

```
showClass("lmerUvcov")
```

maize.G	<i>Genomic relationship matrix for maize lines</i>
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Description

A matrix with relationship between individuals for parents of two heterotic groups. The matrix was computed using 511 SNPs using the function A.mat included in rrBLUP package (Endelman, 2011). The row names and column names of this matrix corresponds to the genotype identifiers for Parent 1 and Parent 2.

maize.Pheno	<i>Phenotypical data for the maize dataset</i>
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Description

Phenotypic data for grain yield and plant height for 100 out of 400 possible crosses originated from 40 inbreed lines belonging to two heterotic groups, 20 lines in each. The data are stored in a data.frame with 6 columns: Location, GCA1 (Parent 1), CGA2 (Parent 2), SCA (hybrid), Yield and PlantHeigh. Records with missing values in the last two columns corresponds to hybrids that were not evaluated in the field and that we need to predict.

ranefUvcov

Extract the conditional means of the random effects

Description

A function to extract the conditional means of the random effects from a fitted model object.

Usage

```
ranefUvcov(object, postVar = FALSE, drop = FALSE, which1 = names(ans), ...)
```

Arguments

object	is an object returned by lmerUvcov.
postVar	a logical argument indicating if the conditional variance-covariance matrices of the random effects should be added as an attribute.
drop	should components of the return value that would be data frames with a single column, usually a column called “(Intercept)”, be returned as named vectors instead?.
which1	character vector of names of grouping factors for which the random effects should be returned.
...	some methods for these generic functions require additional arguments.

Details

The function ranef extract the conditional means for the liner mixed effects model:

$$\mathbf{y} = \mathbf{X}\boldsymbol{\beta} + \mathbf{Z}_1^* \mathbf{u}_1^* + \cdots + \mathbf{Z}_q^* \mathbf{u}_q^* + \mathbf{e},$$

where $\mathbf{Z}_j^* = \mathbf{Z}_j \times \mathbf{L}_j$, with $\mathbf{L}_j$ from Cholesky factorization for $\mathbf{K}_j$. Alternatively, $\mathbf{Z}_j^* = \mathbf{Z}_j \times \boldsymbol{\Gamma}_j \boldsymbol{\Lambda}_j^{1/2}$, with $\boldsymbol{\Gamma}_j$ and $\boldsymbol{\Lambda}_j$ the matrix of eigen-vectors and eigen-values obtained from the eigen-value decomposition for $\mathbf{K}_j$. So, the conditional means of the random effects in the linear mixed effects model:

$$\mathbf{y} = \mathbf{X}\boldsymbol{\beta} + \mathbf{Z}_1 \mathbf{u}_1 + \cdots + \mathbf{Z}_q \mathbf{u}_q + \mathbf{e},$$

are obtained as follows: $\hat{\mathbf{u}}_j = \mathbf{L}_j \hat{\mathbf{u}}_j^*$ if the Cholesky factorization is used or $\hat{\mathbf{u}}_j = \boldsymbol{\Gamma}_j \boldsymbol{\Lambda}_j^{1/2} \hat{\mathbf{u}}_j^*$ if the eigen-value decomposition is used.

Value

A list of data frames, one for each grouping factor.

Author(s)

Paulino Perez-Rodriguez

References

Caamal-Pat D., P. Perez-Rodriguez, J. Crossa, C. Velasco-Cruz, S. Perez-Elizalde, M. Vazquez-Pena. 2021. lme4GS: An R-Package for Genomic Selection. *Front. Genet.* **12**:680569. doi: 10.3389/fgene.2021.680569 doi: 10.3389/fgene.2021.680569

Examples

```
library(BGLR)
library(lme4GS)

#####
#Example wheat
#####
data(wheat)
X<-wheat.X
Z<-scale(X,center=TRUE,scale=TRUE)
G<-tcrossprod(Z)/ncol(Z)
A<-wheat.A
rownames(G)<-colnames(G)<-rownames(A)
y<-wheat.Y[,1]

data<-data.frame(y=y,m_id=rownames(G),a_id=rownames(A))

fm1<-lmerUvcov(y~(1|m_id)+(1|a_id),data=data,
               Uvcov=list(m_id=list(K=G),a_id=list(K=A)))

summary(fm1)

#Predictions
plot(y,predict(fm1))

#Random effects
ranef(fm1)

#Equivalently
ranefUvcov(fm1)
```

ranefUvcovNew

Obtain BLUPs for new levels of random effects with user specified variance covariance-matrices.

Description

Obtain BLUPs for new levels of random effects with user specified variance covariance-matrices.

Usage

```
ranefUvcovNew(object,Uvcov)
```

Arguments

object is an object returned by lmerUvcov.
Uvcov two level list with ids to be predicted and variance covariance matrix that contains information of these ids and the ids used to fit the model.

Details

Assume that the random effect $\mathbf{u}_j \sim N(\mathbf{0}, \sigma_j^2 \mathbf{K}_j)$ and the matrix $\mathbf{K}_j$ is partitioned as follows:

$$\mathbf{u}_j = \begin{pmatrix} \mathbf{u}_{j1} \\ \mathbf{u}_{j2} \end{pmatrix}$$

and

$$\mathbf{K}_j = \begin{bmatrix} \mathbf{K}_{j11} & \mathbf{K}_{j12} \\ \mathbf{K}_{j21} & \mathbf{K}_{j22} \end{bmatrix}$$

The BLUP for $\mathbf{u}_{j2}$ can be obtained as:

$$\mathbf{E}(\mathbf{u}_{j2}|\mathbf{y}_1) = \mathbf{K}_{j21} \mathbf{K}_{j11}^{-1} \hat{\mathbf{u}}_{j1}$$

Value

A list of matrix arrays one for each grouping factor

Author(s)

Paulino Perez-Rodriguez

References

Caamal-Pat D., P. Perez-Rodriguez, J. Crossa, C. Velasco-Cruz, S. Perez-Elizalde, M. Vazquez-Pena. 2021. lme4GS: An R-Package for Genomic Selection. *Front. Genet.* **12**:680569. doi: 10.3389/fgene.2021.680569 doi: 10.3389/fgene.2021.680569

Examples

```
library(BGLR)
library(lme4GS)

data(wheat)
X<-wheat.X
Z<-scale(X,center=TRUE,scale=TRUE)
G<-tcrossprod(Z)/ncol(Z)
```



```

A<-wheat.A
rownames(G)<-colnames(G)<-rownames(A)
y<-wheat.Y[,1]

#Predict 10/100 of records selected at random.
#The data were partitioned in 10 groups at random
#and we predict individuals in group 2.

fold<-2
y_trn<-y[wheat.sets!=fold]
y_tst<-y[wheat.sets==fold]

A_trn=A[wheat.sets!=fold,wheat.sets!=fold]
G_trn=G[wheat.sets!=fold,wheat.sets!=fold]

pheno_trn=data.frame(y_trn=y_trn,m_id=rownames(A_trn),a_id=rownames(G_trn))

#####
#Marker based prediction
#####

fm1<-lmerUvcov(y_trn~1+(1|m_id),data=pheno_trn,Uvcov=list(m_id=list(K=G_trn)))

plot(pheno_trn$y_trn,predict(fm1),xlab="Phenotype",ylab="Pred. Gen. Value")

#BLUP for individuals in the testing set
blup_tst<-ranefUvcovNew(fm1,Uvcov=list(m_id=list(K=G)))
blup_tst<-blup_tst$m_id[,1]

#Comparison
#Check the names
names(y_tst)<-rownames(G)[wheat.sets==fold]
blup_tst<-blup_tst[match(names(y_tst),names(blup_tst))]

yHat_tst<-fixef(fm1)[1]+blup_tst
points(y_tst,yHat_tst,col="red",pch=19)

#Correlation in testing set
cor(y_tst,yHat_tst)

#####
#Pedigree based prediction
#####

fm2<-lmerUvcov(y_trn~1+(1|a_id),data=pheno_trn,Uvcov=list(a_id=list(K=A_trn)))

plot(pheno_trn$y_trn,predict(fm2),xlab="Phenotype",ylab="Pred. Gen. Value")

#BLUP for individuals in the testing set
blup_tst<-ranefUvcovNew(fm2,Uvcov=list(a_id=list(K=A)))
blup_tst<-blup_tst$a_id[,1]

#Comparison

```

```

#Check the names
names(y_tst)<-rownames(A)[wheat.sets==fold]
blup_tst<-blup_tst[match(names(y_tst),names(blup_tst))]

yHat_tst<-fixef(fm2)[1]+blup_tst
points(y_tst,yHat_tst,col="red",pch=19)

#Correlation in testing set
cor(y_tst,yHat_tst)

#####
#Markers + Pedigree based prediction

fm3<-lmerUvcov(y_trn~1+(1|m_id)+(1|a_id),data=pheno_trn,
               Uvcov=list(m_id=list(K=G_trn),a_id=list(K=A_trn)))

plot(pheno_trn$y_trn,predict(fm3),xlab="Phenotype",ylab="Pred. Gen. Value")

#BLUP for individuals in the testing set
blup_tst<-ranefUvcovNew(fm3,Uvcov=list(m_id=list(K=G),a_id=list(K=A)))

blup_tst_m<-blup_tst$m_id[,1]
blup_tst_a<-blup_tst$a_id[,1]

#Comparison
#Check the names
names(y_tst)<-rownames(A)[wheat.sets==fold]
blup_tst_m<-blup_tst_m[match(names(y_tst),names(blup_tst_m))]
blup_tst_a<-blup_tst_a[match(names(y_tst),names(blup_tst_a))]

yHat_tst<-fixef(fm3)[1] + blup_tst_m + blup_tst_a
points(y_tst,yHat_tst,col="red",pch=19)

#Correlation in testing set
cor(y_tst,yHat_tst)

```

theta_optim

Selection of bandwidth parameter for Gaussian and exponential kernels.

Description

Obtain the optimal value of the bandwidth parameter for the Gaussian and exponential kernels.

Usage

```
theta_optim(formula, data = NULL, Uvcov = NULL,
```

```
kernel = list(D = NULL, kernel_type = "gaussian",
              theta_seq = NULL, MRK = NULL),
verbose_lmer= 0L, verbose_grid_search=0L)
```

Arguments

formula	a two-sided linear formula object describing both the fixed-effects and random-effects part of the model, with the response on the left of a '~' operator and the terms, separated by '+' operators, on the right. Random-effects terms are distinguished by vertical bars ' ' separating expressions for design matrices from grouping factors.
data	an optional data frame containing the variables named in 'formula'.
Uvcov	list.
kernel	list with the following elements, i)D: Distance matrix (can be NULL), ii) kernel_type: character, can be either "gaussian" or "exponential", ii)theta_seq: sequence of values for theta from which we select the optimum (can be NULL), iv) MRK: marker matrix from which Euclidean distance is computed (can be NULL).
verbose_lmer	integer scalar, verbose output from optimizeLmer function?. If '> 0' verbose output is generated during the optimization of the parameter estimates, default value is 0L.
verbose_grid_search	integer scalar, if '>0' verbose output is generated, default value is 0L.

Value

A list that contains:

LL	Log-likelihood.
LL.max	Maximum of likelihood.
theta	Sequence of values for the bandwidth.
theta.max	Value of bandwidth when log-likelihood attains the maximum.
fm	Fitted model with the optimum bandwidth parameter.
K.opt	The kernel for the optimum bandwidth parameter.

Author(s)

Paulino Perez-Rodriguez, Diana Caamal-Pat

References

Caamal-Pat D., P. Perez-Rodriguez, J. Crossa, C. Velasco-Cruz, S. Perez-Elizalde, M. Vazquez-Pena. 2021. lme4GS: An R-Package for Genomic Selection. *Front. Genet.* **12**:680569. doi: 10.3389/fgene.2021.680569 doi: 10.3389/fgene.2021.680569

Examples

```
library(BGLR)
library(lme4GS)

data(wheat)

y = wheat.Y[,1]
X = wheat.X
A = wheat.A

rownames(X) <- rownames(A)

#model y=1*mu+Z_1*u_1+e, u_1~NM(0, \sigma_1*KG), KG: Gaussian kernel
wheat = data.frame(y=y, k_id=rownames(X))

fm1 <- theta_optim(y~(1|k_id), data = wheat, Uvcov = NULL,
                  kernel = list(D = NULL, kernel_type = "gaussian",
                                theta_seq = seq(3,8,length.out=10), MRK = X),
                  verbose_lmer=0L,verbose_grid_search=1L)

plot(fm1$theta,fm1$LL,xlab=expression(theta),ylab="Log-Likelihood")
fm1$theta.max
fm1$LL.max
```

wheat.Pedigree	<i>Pedigree infomation for 599 wheat lines</i>
----------------	--

Description

A data.frame with 3 columns: gpid1 and gpid2 which corresponds to the GID of parents 1 and 2 respectively, progenie which corresponds to the GIDs of progenie.

wheat.Pheno	<i>Phenotypical data for the wheat dataset</i>
-------------	--

Description

A data.frame with 4 columns, Env for environments, Rep for replicates, GID for genotype identifiers, Yield for grain yield.

wheat.sets	<i>Sets for cross validation (CV)</i>
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Description

Is a vector (599 x 1) that assigns observations to 10 disjoint sets; the assignment was generated at random. This is used later to conduct a 10-fold CV.

Source

International Maize and Wheat Improvement Center (CIMMYT), Mexico.

wheat.X	<i>Molecular markers</i>
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Description

Is a matrix (599 x 1279) with DArT genotypes; data are from pure lines and genotypes were coded as 0/1 denoting the absence/presence of the DArT. Markers with a minor allele frequency lower than 0.05 were removed, and missing genotypes were imputed with samples from the marginal distribution of marker genotypes, that is, $x_{ij} = \text{Bernoulli}(\hat{p}_j)$, where $\hat{p}_j$ is the estimated allele frequency computed from the non-missing genotypes. The number of DArT MMs after edition was 1279.

Source

International Maize and Wheat Improvement Center (CIMMYT), Mexico.

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